There's no harm in having too much: A comprehensive toolbox of methods in trophic ecology

Nabil Majdi a,b,c,1, Nicolas Hette-Tronquart a,d,l,q,1, Etienne Auclair a,e, Alexandre Bec a,f, Tiphaine Chouvelon a,g, Bruno Cognie a,i, Michael Danger a,h, Priscilla Decottignies a,i, Aurélie Dessier a,i, Christian Desvillettes a,f, Stanislas Dubois a,k, Christine Dupuy a,i, Clémentine Fritsch a,i, Cédric Gauchere l a,m, Mickaël Hedde a,n, Franck Jabot a,o, Sebastien Lefebvre a,p,q, Martin P. Marzloff a,k, Benjamin Pey c, Nathalie Peyraud a,e, Thibaut Powolny a,i, Régis Sabbadin a,e, Elisa Thébault a,r, Marie-Elodie Perga a,s

⁎ Corresponding author at: GRET (Groupe de Recherche en Ecologie Trophique), GDR 3716 CNRS INEE INRA, France.

1 The authors contributed equally to this paper.

Trophic ecology is the study of feeding interactions and food acquisition by organisms. It includes the causes and consequences of those behaviours at all levels of biological organisation. As a field of research, it crosses many disciplinary boundaries and provides knowledge that is pertinent to many other areas of ecology. Here we list and categorise the methods available to trophic ecologists whose toolbox has broadened considerably in recent years. They encompass empirical and numerical approaches with focus ranging from molecules to ecosystems. We further examine the relationship of each method to features such as the scale of observation (from microbes to largest organisms) and organisational level (from individuals to ecosystems) as well as the ecological question the method is capable of answering (from detecting predator-prey relationships to studying implications and consequences at different scales). Our survey reveals a very wide range of methodologies, each more-or-less appropriate for a particular line of research. It also identifies deficits, for example, trophic interactions at microscopic scales, for which empirical methods have hardly been used, as well as trophic models that have failed to consider fluxes at the ecosystem scale. Furthermore, we note that the combination of methodologies remains under-exploited due to great opportunities to solve complex ecological questions and to foster the emergence of new insights and hypotheses regarding organism, population and/or ecosystem properties.

© 2018 Elsevier Inc. All rights reserved.
1. Introduction

Trophic ecology covers the ecological implications of feeding. As such, it embraces the act of food acquisition and its consequences at the individual, population and community levels up to functional outputs at the scale of the whole ecosystem. The field of trophic ecology, in which the concept of food web is embedded, is broader than organismal detection and the description of inter-species feeding relationships. Briefly, at the smallest organisational levels, feeding is intimately linked to nutrition and ecophysiology, as the quantity and quality of mineral or organic food resources regulate individual survival, growth and fecundity with upscaling effects on population demography (Elser et al., 1996). Except for lithotrophic and saprophytic organisms, food acquisition also implies that feeding interactions generate reciprocal effects on predator and prey demography (Lotka, 1925; Volterra, 1926). Trophic relationships are also responsible for species inter-dependencies and contribute to shaping behaviours, driving phenotype selection and eco-evolutionary feedbacks (Post and Palkovacs, 2009; Clinchy et al., 2013). A trophic transition can also be biogeochromatically regarded as a way to transfer mineral or organic matter between two trophic levels with more-or-less efficiency (“trophic efficiency concept” of Lindeman, 1942, “ecological stoichiometry” see e.g. Elser et al., 1996), or transformations in its biochemical quality (“trophic upgrading”; Klein-Breteler et al., 1999). The concept of food web brings trophic ecology to the community and ecosystem scales. Food webs allow exploring the network of consumer-resource interactions, considering its assemblage rules and interaction strength and the consequences of network structure to overall community composition and stability, up to emergent properties of the ecosystem — like carbon fixation (see Layman et al., 2015, and references therein for a synthesis of major concept achievements in food web ecology).

The broadness of trophic ecology implies a diversity of methodologies, along with considerable improvements and developments both in field and laboratory methods. In addition, numerical approaches have flourished and now support trophic ecologists in their efforts to conceptualise their theories and to modify them based on empirical data (e.g. Proulx et al., 2005; D’Alelio et al., 2016).

Considering this large and fast-evolving bouquet of methodologies, we felt that an up-to-date overview of existing methods, merging both empirical and numerical technics, has been so far missing. Here, by providing a technical overview, we hope to facilitate both upstream decision-making and the inclusion of a broader diversity of research paths within otherwise well-defined methodological road maps. Nevertheless, we do not pretend to provide either an exhaustive critical review of every method relevant to trophic ecology or systematic pairwise comparisons of all methods. For this, we refer readers to the comprehensive book by Garvey and Whiles (2017) and to more specific methodological reviews referenced hereafter. Because the choice of a method primarily depends on its ability to answer a question at hand, we have structured this review accordingly, grouping the methodological reviews referenced hereafter. Never-
2. Observations of feeding activity

2.1. Observations of feeding activity

The most straightforward and oldest approach to study feeding interactions relies on observations. Feeding observations can be made with the naked eye and simple equipment (e.g. binoculars) or using sophisticated, miniaturised, high-magnification optical devices, including remote underwater video stations, camera traps and drones (e.g. Linchant et al., 2015). This allows large-scale records of feeding behaviours in remote ecosystems that are poorly studied. In addition, animal-borne video systems offer the animal’s perspective (e.g. Moll et al., 2007). For microscopic or suspension-feeding animals, most observations are restricted to laboratory conditions, for example using a camera attached to a microscope (e.g. Dupuy et al., 2010) or by in vivo video endoscopy (e.g. Prasetiya et al., 2017). To complement direct observations, an emerging approach consists in remotely monitoring multiple biological and environmental factors that constitute surrogates for feeding activity, such as the movements of body parts, oesophageal or gastric temperatures and geographical positions, using animal-attached bio-logging devices (Ropert-Coudert and Wilson, 2005). This should further progress with the transfer of geo-referenced human mobility data analysis techniques to animals (Thums et al., 2018). If direct observation of feeding activity is impracticable, food intake can be inferred from the disappearance of food items. For example, suspension-feeder activity is most commonly assessed using food-clearance experiments or flow-through chamber methods (see e.g. Ward and Shumway, 2004). Different parameters can be obtained this way, such as clearance rate, retention and sorting efficiencies, ingestion, and egestion rates.

2.2. Gut content and faeces analysis

Observing the food remains in digestive tracts of animals is the most common approach to get a basic, timely knowledge of predator–prey relationships. Some studies have used massive gut contents datasets to evaluate and compare trophic networks in different ecosystems (e.g. Winemiller, 1990). Especially in fishes, typical methodologies and indices have been developed, and some like the percentage of prey occurrence may emerge as relatively reliable proxies of diet composition (see Baker et al., 2014). Rejection pellets and faeces can also be collected and inspected for undigested prey items (e.g. Kusmer, 1990). In most cases, a dietary assessment is based on the recovery of hard/recalcitrant prey parts (e.g. bones, fur, diatom frustules), which makes a reference collection of such parts often necessary to identify the respective prey organism.

The main advantages of this type of dietary analysis consist in: (1) being accessible, (2) being applicable at relatively large spatial and temporal scales and (3) enabling assessments of prey size (e.g. measuring otoliths or vertebrae). Among the main drawbacks are (1) the discrepancies in taxonomic distinctness (as the ingested items can be identified either at the species level or at a higher taxonomic level), (2) the inability to account for labile food items and (3) the requirement of comprehensive taxonomic expertise. (4) Ethical issues are also raised by this approach, particularly when animals must be dissected. Alternative techniques that avoid dissection can be used to recover stomach contents, such as stomach flushing or regurgitation (e.g. Pierce and Boyle, 1991; Legler and Sullivan, 1979; Duffy and Jackson, 1986). In addition, as many microscopic animals have transparent bodies, their gut contents can often be examined under the microscope without dissection. For example, confocal laser scanning microscopy can be used to detect the auto-fluorescence of micro-algal pigments in the gut of rotifers (Mialet et al., 2013), and chromatography can serve to quantify ingested biomarker pigments extracted from guts (e.g. Majdi et al., 2012).

Molecular approaches can also be used to decipher ingested prey remains. It is based on the use of specific prey primers to amplify the prey DNA contained in the gut or faeces of the predator, followed by (meta) barcoding (for a review see King et al., 2008). The use of molecular gut content has really expanded in recent years and has proved useful to assess, for instance, the diet of minute animals such as soil micro- and meso-arthropods (e.g. Eitzinger et al., 2013; Heidemann et al., 2014), and zooplankton (e.g. Ho et al., 2017). The major advantages of analysing gut content DNA are: (1) high taxonomic resolution without taxonomic expertise, (2) the inclusion of partly digested, unidentifiable prey chunks and, potentially, (3) the acquisition of data on the microbial assemblages ingested or persisting in the gut (i.e. gut microbiomes, see, e.g. Derycke et al., 2016). However, besides substantial costs, there are several disadvantages inherent to molecular approaches, especially (1) high risk of sample contamination and false positives, (2) need for specific primers to identify uncommon prey taxa, (3) absence in public gene repositories of sequence data for the DNA of numerous prey types, (4) bias in quantitative extrapolations due to the differential timing of DNA degradation during digestion or the specificity of certain DNA extraction protocols and (5) the inability to detect cannibalistic behaviour (Traugott et al., 2013).

2.3. Parasites as biological tags

Since some parasites are transmitted only through food ingestion, they serve as evidence of a feeding interaction and indicate that the host species has fed in geographical areas where the prevalence of this parasite is high. Similarly, accurate information about the feeding grounds and past movements of animal hosts can be surmised based on the presence of multiple parasite species differing in their endemic areas (MacKenzie and Abaunza, 1998). Yet, to be reliable biological tags, parasites must have: (1) distinct prevalences in different studied areas, (2) limited inter-annual or seasonal variations in those prevalences, (3) a high specificity for the microhabitat within the host’s body, (4) a lifespan fitting the aim/scale of the study, (5) no strong pathological effects on the host and (6) preferably, a direct and single-host life cycle (facilitating interpretations). They must also be readily detectable and identifiable. Helminths generally fulfill all of these criteria and are thus the most commonly used trophic tracers (see MacKenzie, 2002).

The use of parasites as trophic tracers has thus far been limited to marine ecosystems, but, given the ubiquity of parasites (Marcogliese, 2004), implementation of this method in other ecosystems is certainly possible (see Holmstad et al., 2004). Additional advantages of parasites are that they provide information even when their host’s stomach is empty (Baldwin et al., 2008) and indicate long-term feeding, as they can accumulate in their hosts for months or longer (Marcogliese and Cone, 1997). However, this method also requires that the animals are killed and dissected, the method is time-consuming and considerable taxonomic expertise is required. Some of these shortcomings might be overcome using DNA barcoding, which is inexpensive (~SE/parasite), or by coupling behavioural and molecular approaches (Poulin, 2010). Anyway, the use of a parasite as a biological tag relies on an at least partial understanding of the parasite’s life cycle, which probably explains why they are mostly used in fish species so far.
2.4. Tracers of biomass

Trophic tracers of biomass origin are elements or molecules that an organism can gain only from its trophic resources. The presence of a tracer in the biomass of an organism indicates the consumption and assimilation of a resource containing that tracer, as well as whether and how much the different trophic resources have contributed to the biomass of the organism (or its targeted tissue), provided that the potential resources differ in their tracer composition. Moreover, the presence of a tracer not only reflects the consumption of a resource, it can also provide information on how the tracer is retained and transferred within the consumer, such as by selective digestion or assimilation (Gannes et al., 1997) or routing to different tissues (Carleton et al., 2008).

2.4.1. Stable isotope composition of bulk tissues

One of the most common trophic-tracer–based methods is the determination of the stable isotope composition of bulk tissues, such as muscle, blood, liver, feather, hair, fin, or whole organism. In trophic ecology, C, N and S are the tracers most frequently used in stable isotopic analysis (SIA) (Fry, 2006), but H/D and O isotopes are also employed (Vander Zanden et al., 2016). The method is based on predictable differences (trophic discrimination factors, TDFs) between the isotopic composition of an organism and that of its food resources. For C and S, the difference is limited and these elements are mainly used to trace the origins of the food resources from which the consumers’ biomass has been synthesized (Boecklen et al., 2011), whereas the TDF for N is larger, allowing its use in estimates of trophic position (Post, 2002). However, the variability in the TDFs depending on environments, trophic levels, taxa, tissues, etc. (Bond and Diamond, 2011) is a potential confounding factor that has been highlighted in several studies (e.g. Bastos et al., 2017). In addition, the high variability of stable isotope compositions in space and time (e.g. Hyndes et al., 2013) must also be considered. Several tools are used to analyse stable isotope data. Their most recent versions generally adopt a similar Bayesian approach to account for the sources of uncertainty (e.g. Parnell et al., 2013) and include associated confidence intervals. Bayesian approaches also allow introducing informative priors, increasing the accuracy and precision of the estimates. We distinguished three main types of tools: (1) those devoted to the estimation of trophic position (e.g. TrophicPosition, Quezada-Romegialli et al., 2018), (2) isotopic metrics to measure specimen or species positions in the isotopic space and to compare the variability of isotopic signatures across individuals, populations and communities (SIBER, Jackson et al., 2011; Cucherousset and Villéger, 2015) and (3) mixing models to estimate the contribution of each potential trophic resource to the biomass of an organism (see Philips, 2012 for a review). The relevance of SIA-derived results is thus strongly tied to isotopic discrimination between resources, the spatial and temporal variability of the isotopic compositions captured by resource/consumer sampling, and an accurate estimate of the TDF. SIA is not well suited to situations involving large numbers of potential resources (e.g. Robinson et al., 2018). The integration of the food isotopic signal into the consumer’s tissue depends on the rate at which tissue are renewed by somatic growth or metabolic tissue replacement (which determines isotopic turnover). Issues related to isotopic turnover thereby limit the relevance of the method for individuals (i.e. adults) or seasons of reduced somatic growth (gonadic growth or maintenance metabolism, Perga and Gerdeaux, 2005). Issues related to differential isotopic routing (Gannes et al., 1998; Del Rio et al., 2009) and unequal TDF in plant versus animal food sources (Perga and Grey, 2010) also limit the detection of omnivory for individuals and species exploiting both plant and animal resources. As a result, SIA enables quantitative estimates of elemental fluxes within the different steps and habitats of a food web, but it is a less than ideal method with which to assess the diversity and specificity of a consumer’s diet.

When natural isotope abundance does not provide the necessary discrimination between sources, for example, to overcome uncertainties linked to TDF variability, small quantities of isotopically labelled sources (stable or radioactive) can be injected in- or ex-situ to track C and/or N pathways. This method of isotope labelling, or Stable Isotope Probing (SIP) aims to track the integration of the labelled elements within food webs (Pace et al., 2007). Primary producers can be selectively labelled through addition of 13C-enriched bicarbonate/carbon dioxide or 15N-enriched ammonium or nitrate directly in situ (see e.g. Middelburg et al., 2000) to track the fate of primary production through food webs. Similarly, SIP based on dissolved organic matter (e.g. Hall and Meyer, 1998), phytodetritus (e.g. Sarnthein et al., 2014) or animal faeces (e.g. Dunjait et al., 2009) are used to label microbial decomposers and the fate of dead organic matter through “brown” food webs. The mode (single addition, pulse chase, or constant levels) and the duration (a few hours to several months) at which the label is introduced into the system determine the time and scale at which the fate of the labelled substrate is tracked (from microbial interactions up to the whole ecosystem), and eventually to document the dynamics of the processes (when the pulse chase mode is adopted) (e.g. Maxfield et al., 2012). If studies using isotope labelling are relieved from the issues of TDF uncertainties encountered for SIA in natural abundances, they also suffer for issues related to isotope routing. Indeed, structural components of plant material behave differently than metabolic components in terms of microbial utilisation, grazing, and assimilation (see e.g. Soong et al., 2014). The success of isotope-labelling experiments is thereby strongly tied to the ability to uniformly label throughout the food source, or instead to differentially label the structural and metabolic components of the source (Soong et al., 2014). SIP studies have proven powerful in documenting microbial involvement in biogeochemical processes in soils and sediments and in quantifying the trophic links between the microbial and macro-faunal worlds (e.g. Crotty et al., 2012; Middelburg, 2014).

2.4.2. Fatty acid trophic biomarkers

Since the early 1980s, fatty acids (FAs) have been widely used to study predator–prey relationships and to trace the origin and transfer of organic matter in food webs (St-John and Lund, 1996). The use of FAs as trophic markers relies on the specific FA patterns exhibited by basal resources that are transferred to and identifiable in consumers. As such, FAs have been analysed in many species to infer feeding relationships in aquatic environments for example (Arts et al., 2009). They are also increasingly used to shed light on feeding channels in soil environments (e.g. Russel et al., 2007). Most often, this method requires tracing specific FAs of prey that cannot be synthesized de novo by the targeted community of consumers/predators (e.g. 16:4ω3 for chlorophytes, 16:4ω4 for diatoms, 22:1ω11 for Calanoid copepods). The detection of those prey-specific FAs in the tissues of consumers/predators thus reveals the assimilation pathway of the prey. However, many fatty acids (e.g. 16:0, 18:1ω9c, 16:0ω3c) occur in a wide range of organisms restraining the accuracy of the approach (Dalsgaard et al., 2003). For this matter, the comparison of FA ratios can improve the interpretation of results (Parrish, 2013). For example tracking the 16:1ω7/16:0 ω6 ratio for diatoms or the 22:6ω3/20:5ω3 ratio for chrysophytes can be used to demonstrate that these microalgae are assimilated by zooplankton micromucrustaceans (Desvillettes et al., 1997; Pepin et al., 2011). Indeed, the reliability of FA-based trophic markers requires in-depth knowledge of the lipid metabolism of both primary producers and the targeted animal species as a prerequisite. However, while these metabolic processes may obscure the identification of actual food resources, FA-based methods generally provide sufficient information on foraging patterns.

To date, most studies using FA have been qualitative with the presence/absence of a source-specific FA being used as an indicator of the existence of a trophic interaction. A Quantitative FA Signature Analysis (QFASA) has been more recently developed to enable the estimation of the proportions of different sources in a predator’s diet (Iversen et al., 2004). Basically, the FA signatures of consumers are modelled as a mixture of the FA signatures of major food resources, which provides
a quantitative estimate of the proportions of those resources. The method is based on the calculation of the statistical distance between predator FA signatures and mean prey FA signatures. The predator’s FA patterns are corrected according to the degree of modification linked to its FA metabolism. Calibration coefficients are then necessary and calculated using experimental diet studies. These studies are designed to assess the selective retention of dietary fatty acids, to detect fatty acid bioconversion or de novo synthesis. The acceptance of this method by researchers studying the trophic ecology of marine mammals for example will likely be broadened by the recent development of a dedicated R-package (Bromaghin, 2017). According to Happel et al. (2016), however, as for mixing models in SIA, the use of QFAS should be limited to the predators that consume a small number of species and that can be studied experimentally as well as in the wild.

2.4.3. Compound specific isotopic analyses (CSIA)

Combining detection of biomarkers (lipids and amino-acids mainly, but also amino-sugars) to stable isotope analyses, CSIA is able to trace the sources and pathways of numerous natural as well as anthropogenic organic compounds. In ecology, first utilizations of CSIA focused on chemotaxonomic markers of microorganisms. Although the role of microorganisms is crucial in processing and transferring organic matter in food webs, their isotopic signatures were rarely reported due to the difficulty of isolating specific microbial biomass from natural samples. Using $^{13}$C labelled substrates Boschker et al. (1998) described a CSIA approach to directly link specific microbial processes with the organism involved, based on the stable carbon isotope labelling of individual lipid biomarkers. In the same way, Werne et al. (2002) followed the $^{13}$C depleted signal of methane in cold seep sediment through different specific microbial markers and could trace with smart elegance the flow of methane-derived-carbon through anaerobic methane-oxidizing archaea (archaeol and sn-3-hydroxyarchaeol) into sulfate-reducing bacteria (C15 and C17 iso and anteiso fatty acids, diakyl glycerol diethers), as well as into aerobic methanone-oxidizing bacteria (diploptene, diploptol) up to bacterivorous ciliates (tetrhymanol). Stable isotope probing of amino-sugars is regarded as a promising tool to investigate fungal-bacterial interactions in soils (Bode et al., 2013). Thus, CSIA can be used successfully for elucidating source and fate of naturally or artificially labelled substrates.

In other cases, the isotopic fractionation between substrates and chemotaxonomic markers must be known. Food webs studies have thus applied CSIA on essential lipid compounds such as polyunsaturated fatty acids (PUFAs) or sterols by assuming that TDFs would be negligible. However, an unpredictable isotopic fractionation ranging from 0% to 4% still occurs during trophic transfer of essential lipid compounds such sterols (Chamberlain et al., 2004) or PUFAs (Bec et al., 2011; Gladyshev et al., 2016). Moreover, the intermolecular variability of $\delta^{13}$C values within the same sample may be relatively high, thereby challenging again data interpretation. For example, the difference between fatty acids $\delta^{13}$C values within a single cultivated alga may be up to 7.6‰ (Schouten et al., 1998). Thus, interpretation of CSIA data could be constrained in case the potential endmembers are not sufficiently isotopically different. However, when the conditions are met, CSIA could trace limiting compounds and eventually detect minor food sources unseen by classical methods. For example, CSIA has revealed an uncoupling between essential compounds and major organic matter transfers highlighting functional importance of minor food sources of great nutritional importance (Koussoroplis et al., 2010). An increasing attention is now being paid to amino acids-CSIA since McClelland and Montoya (2002) showed that the $^{15}$N values of some amino acids such as phenylalanine in consumers are very similar to those in the producers while the $^{15}$N values of other amino acids such as glutamic acid become enriched in $^{15}$N with each trophic transfer. Thus, due to this differential $^{15}$N-enrichment with trophic transfer, $\delta^{15}$N of amino acids in a consumer provide a measurement of the $\delta^{15}$N baseline as well as its trophic position.

Coupled with stable isotopic probing, ribosomal RNA or DNA can also be used as integrative tracers. $^{13}$C- or $^{15}$N-labelled RNA or DNA are heavier so that they can be separated from unlabelled genetic material using isopycnic centrifugation gradients. Further amplification and barcoding of the isolated fractions makes possible to identify the organism associated with the assimilation of a specific resource or organic contaminant (Neufeld et al., 2007). In microbial ecology, this is a promising approach to detect specific functional guilds of microbes in complex environmental samples (e.g. methanotrophic bacteria), but it can also be scaled-up to other consumers, such as fungi and flagellates, to reveal matter flows in microbial food webs (Lueders et al., 2003).

2.4.4. Contaminants as trophic tracers

Both trace metals (inorganic elements) and xenobiotic organic compounds can fulfil the role of trophic tracers (Ramos and González-Solis, 2011) when these contaminants enter trophic pathways and differentially bio-accumulate in food sources or prey and consequently in consumers or top predators (e.g. Walters et al., 2008; Pitt et al., 2017). Selected contaminants can thus be used to infer trophic relationships, including the prey preferences of consumers/predators, foraging areas or food webs exploited by different individuals or populations (e.g. Larsson et al., 1990; Deshpande et al., 2016) and even trophic positions and organic matter flux when bio-magnification occurs (Kelly et al., 2007). Therefore, these indirect tracers can be particularly helpful when there is inherent difficulty to directly observe trophic relationships (e.g. for marine species). Another advantage of using contami-nants in a trophic perspective is that it also informs on the contamination status of the species and food webs of concern, which may be very interesting for ecotoxicological and/or conservation biology perspectives as well. For example, a survey of cadmium (Cd) in common dolphins from the North-East Atlantic allowed neric and oceanic populations to be distinguished based on the Cd concentrations in their kidneys (long-term accumulation organ in mammals) and their distinct preferential prey (Lahaye et al., 2005). Cephalopods and especially oceanic squids (i.e. the preferential prey of oceanic dolphins compared to neric ones) indeed display very high burden of Cd relative to fish prey species, and were thus proved to constitute a significant vector of Cd for their predators (Caurant and Amiard-Triquet, 1995; Bustamante et al., 1998).

However, while monitoring contaminants can yield information on both trophic interactions and matter fluxes, good knowledge of the contaminants’ characteristics (biogeochemical cycles, the distribution and persistence of the contaminant in organisms and the environment), the target organism’s biology, and the structure of food webs are essential. Another prerequisite is an obvious source of the contaminant and its clear partitioning. In fact, contaminants are rarely used alone as trophic tracer tools, and rather complement other intrinsic trophic markers such as C, N, S isotopic ratios or FA profiles (e.g. Krahm et al., 2007; Hebert et al., 2009; Praca et al., 2011). For example, in prey with similar FA but different contaminant profiles, use of the latter allows the discrimination of different diets in the predator (while the use of FA alone did not). Nevertheless, the use of contaminants as trophic tracers requires that the analysed tissues are carefully chosen, because the type of tissue strongly influences the turnover rate of the contaminant-containing compound, which in turn modifies the integration time of the contaminant itself. Finally, like parasites-based approaches, the toxic effects of a contaminant may alter the feeding strategy of the organism. However, as long as the contaminants are able to track these potentially altered foraging behaviours, they fulfill their role as trophic tracers. With the recent development and improvement of modelling tools such as the “Ecotracer” module (Walters and Christensen, 2018) in the Ecopath with Ecosim software package (see Sections 3.3 and 5.1), the potential of contaminants as trophic tracers could be further extended.
3. Connecting species within networks from ecological data (inference methods)

Because establishing complete inventories of trophic interactions within a system is virtually impossible, inference methods can be used to reconstruct trophic networks when the ecological datasets are incomplete (Jordan, 2016). Inference methods are thus specifically useful when empirical approaches (such as those described in Section 2) cannot be implemented to assess trophic interactions. In that case, inference methods combine empirical data about the community composition (mostly occurrence/abundance of the different species) and theoretical knowledge about trophic ecology (e.g. trophic behaviour of the species) to infer the trophic interactions within the community. It is worth noting that inference methods were not specifically developed for trophic interactions, but for all kinds of interactions (from causal relationships among variables, to social networks). There are several ways to infer interactions (e.g. using Gaussian distributions, logical interaction rules or likelihood postulates), each leading to a different method that can be applied to trophic interactions. In the following, we only focus on the inference methods that are mainly used in the reconstruction of trophic networks.

3.1. Graphical models

A Graphical model is a probabilistic model using a graph to represent the dependencies among variables. In the case of trophic networks, the variables are species abundances or occurrences and the edges in the graph can be interpreted as trophic interactions. Generally, the graph structure is given by expert knowledge, and graphical models are used to make predictions about species persistence. Conversely, it is also possible to learn the graph structure, in our case the trophic network, using empirical data. The principle is to draw an edge between two species in the community, whenever their abundances/occurrences are not independent. Two main kinds of graphical models are used in trophic ecology: Gaussian Graphical Models (GGMs) (Højsgaard et al., 2012) and Bayesian Networks (BNs) (Jensen and Nielsen, 2007).

With GGMs, independence between two species is assessed from the empirical data with the precision matrix (i.e. the inverse of the covariance matrix of the model). The resulting network is an undirected interaction network whose edges are weighted by the coefficient of the precision matrix. Expert knowledge is then required to determine the nature and the direction of the inferred interactions (trophic or non-trophic links). In addition, it is often necessary to limit the number of inferred interactions to the most relevant ones. To do so, GGMs are coupled with sparse regularisation techniques, such as the graphical LASSO (for Least Absolute Shrinkage and Selection Operator; Friedman et al., 2008). Faisal et al. (2010) tested different GGM procedures on large-scale spatial data sets describing abundance records for 39 species of European Warblers. They were able to reconstruct in silico the ecological networks, and from the inferred networks, they could compare the relative role of bioclimatic versus biotic interactions on the species spatial distribution.

Unlike GGMs, BNs and their temporally explicit extension, dynamic-BNs (Dean and Kanazawa, 1989) rely on directed graphs. Concretely, it means that an edge can be interpreted directly as a predation link. BNs also differ from GGMs by the learning process of the graph. In the case of BNs, every possible graph is used to assess species abundances/occurrences. Then, each graph receives a score, depending on both the fit between observed and inferred abundances/occurrences and graph complexity. Scores are calculated using different statistical score functions (e.g. Bayesian information criterion), but the underlying principle is the same: to find a graph (in our case a trophic network) that maximises the score, exactly or approximately. In trophic ecology, BNs and dynamic-BNs have been successfully applied to infer feeding interactions among species based on synthetic data (Aderhold et al., 2012), by combining expert knowledge and field data, such as long-term catches per unit effort of fisheries (Trifonova et al., 2015), or based on the presence-absence time series of a fish community (Sander et al., 2017).

3.2. Logic-based approaches

Logic-based approaches differ from graphical models by the way they reconstruct trophic networks. The principle is not based on statistical approaches, but on a set of logical rules (in the form of: “if premise ‘then’ consequence”) that is used to reconstruct trophic networks. The main advantage of such approaches is to establish the rules by combining background knowledge about the species and their environment and empirical data. In trophic ecology, two techniques are commonly used to determine the set of logical rules: Inductive logic programming (Muggleton, 1991) when background knowledge is available; or meta-interpretive learning (Muggleton et al., 2014) when background knowledge is incomplete or lacking, (e.g. when the trophic ecology of the species is not known and is actually the object of the attention). Logic-based approaches have been successfully employed to automatically generate trophic networks. For instance, Bohan et al. (2011) reconstruct the trophic network of invertebrate communities within arable fields in Great Britain, using logic-based machine-learning algorithms, species occurrence data, and background knowledge, such as species body-size and functional groups.

3.3. Linear inverse inference

In linear inverse inference, trophic networks are represented in a steady state, as static webs of model compartments (e.g. species or group of species) that are linked together by linear interactions (Van Oevelen et al., 2010). The approach is particularly suited to infer the magnitude of missing interactions, a very common situation in many ecosystems given the high-dimensionality of trophic networks and the scarcity of information across all trophic levels. Linear inverse inference estimates the missing interactions by ensuring mass-balance across all compartments. Practically, different kinds of techniques can be used (see Van Oevelen et al., 2010) to select the set of parameters that produce the best fit between the modelled abundances (or biomasses) of each compartment and empirical observations. The Ecopath software (see Christensen and Pauly, 1992) constitutes a typical example of this approach. In addition, linear inverse inference can be coupled with ecological stoichiometry or stable isotope analysis to further constrain the solution range of the reconstructed food webs. For instance, Pacella et al. (2013) associated isotope data with linear inverse modelling to reconstruct the feasible food webs within the Marennes-Oléron intertidal seagrass bed.

3.4. Approximate Bayesian computation

Bayesian inference methods rely on the computation of a likelihood, which is the probability density of the observed data under the studied model. It is associated with the parameter values of the model. As a likelihood computation is not always possible, approximate Bayesian methods can be used instead, the most common one being the Approximate Bayesian Computation (ABC). This approach originates from the field of population genetics (Tavaré et al., 1997) but has been successfully applied in trophic ecology to decipher the impacts of trophic interactions on the spatial structure of communities (e.g. Jabot and Bascompte, 2012). In an ABC, the likelihood computation step is replaced by intensive simulations of the model with variable parameter values, so as to select the values that lead to the best match between simulations and empirical data. ABC can be further coupled to efficient algorithms, such as sequential or traditional Markov Chain Monte Carlo methods (Jabot et al., 2013). For example, this approach was used to evidence the presence of density-dependent dispersal in
spatially distributed food webs, as well as its implication for food web spatial structure (Melián et al., 2015).

4. Quantifying the consequences of feeding from individuals to ecosystems

4.1. Ecophysiological markers

Nutritional status and/or body condition can be assessed using diverse types of information, such as foraging success or the energy stored by individuals (Schulte-Hostedde et al., 2001). Using ecophysiological approaches (such as nutritional status or body condition) can be helpful to (1) infer feeding activities and foraging history of individuals and populations, and (2) investigate the consequences of trophic relationships and resource availability on individual health and fitness as well as population and community dynamics. Despite the existence of extremely invasive or lethal methods currently available to assess body condition, body composition, and nutritional status, non-lethal techniques are increasingly used. As described in recent reviews, these techniques include body condition indices, body fluid biochemistry, chemical dilution techniques (isotope dilution: change in concentration of isotopes of hydrogen and/or oxygen in body water, and gas dilution: change in concentration of lipid soluble gas), electrical approaches such as body electrical conductivity and bioelectrical impedance, scanning methods (e.g. magnetic resonance imaging, ultrasound scanning, dual-energy X-ray absorptiometry, computed tomography), molecular analyses (e.g. gene expression), and measurements of stress hormones and immune responses (Stevenson and Woods, 2006; Wilder et al., 2016).

In addition, animal performance (i.e. various life-history traits related to fitness, such as survival, development rate and growth, resistance to starvation, tests of stamina, and reproductive success) can be measured. These data are often used to assess the physical condition of the animal, or to evaluate the “quality” of its food and/or habitat and determining factors of trophic interactions or driver of diet selection (Harrison et al., 2011; Jakob et al., 1996; Raubenheimer and Simpson, 2004; Visanuvimol and Bertram, 2011). Metabolism is also a physiological indicator; it can be determined directly through calorimetry (e.g. Ruiz et al., 2018), and indirectly through respirometry (Sadowska et al., 2008; Urrejola et al., 2011). Among this wide range of techniques, we focus here on those most widely used, easy to conduct, least invasive, and least expensive methods, namely body condition indices and body fluid biochemistry.

4.1.1. Body condition

Body composition, or the amount of fat vs. fat-free body mass, is probably one of the most direct indicators of body condition. It provides a quantitative assessment of energy storage because lipid stores constitute the principal form of energy reserves in animals (Schulte-Hostedde et al., 2001; Walsberg, 1988). Body composition is mostly assessed with two or three compartments: fat mass, lean dry mass/water mass, and protein and ash mass (Jakob et al., 1996; Peig and Green, 2009; Reynolds et al., 2009; Schulte-Hostedde et al., 2005). Alternative, but destructive methods that can be conducted in study designs based on carcass collection for instance include the sampling and weighing of specific subcutaneous fat depots during dissection or the use of supplementary somatic indices such as those of the liver or pancreas (Stevenson and Woods, 2006). However, such methods are tedious, time-consuming, intrusive, as well as destructive.

Consequently, other methods have been proposed to measure body composition or fat stores, notably total body electrical conductivity (Reynolds et al., 2009; Walsberg, 1988), magnetic resonance imaging (McGuire and Guggielmo, 2010), and body composition modelling (Molnar et al., 2009). In certain taxonomic groups (such as birds) subcutaneous adipose depots can be assessed by simple visual inspection (Labocha and Hayes, 2012), whereas others (e.g. small mammals) require simple measurements, such as pelvic circumference considered to be representative of fat content (Labocha et al., 2014). In microscopic organisms, coherent anti-Stokes Raman spectroscopy can be used to non-invasively observe lipid droplets in tissues (e.g. Fueser et al., 2018).

Body condition indices based on morphometrics have also been developed. Most discriminate between the mass of the individual associated with body structural size (body weight) and the mass related to energy reserves (Green, 2001; Peig and Green, 2009; Schulte-Hostedde et al., 2005). Accordingly, the greater the body weight is, the larger are the energy reserves, and the better is the body condition. Body condition indices thus allow the relative size of the body’s energy stores to be computed, taking into account body structural size components. Historically, this was achieved using the residuals of the regression (typically, ordinary least squares regression) between body weight and some index of body size, or by calculating the scaled mass index (Green, 2001; Peig and Green, 2009, 2010; Schulte-Hostedde et al., 2005). Although much debated (Wilder et al., 2016), body condition indices based on morphometrics have been, and are still, widely applied (Labocha et al., 2014; Labocha and Hayes, 2012; Wilder et al., 2016).

4.1.2. Body fluid biochemistry

Several (micro-) nutrients needed to fulfill basic energetic requirements and metabolism cannot be synthesized de novo and can only be acquired from food (Kohl et al., 2015; McWilliams, 2011). These essential trace elements include dietary minerals (e.g. Cu, Co, Fe, Mn, Mo, Se, Zn), carotenoids, vitamins, and some amino acids and FAs (Costantini et al., 2010; Harrison et al., 2011; Kohl et al., 2015; McWilliams, 2011). By measuring the levels of certain metabolites, electrolytes, and enzymes, the depletion of some essential or limiting dietary element, and thus the health state (or the presence of food deficiencies) of the individual, can be determined directly. Moreover, information are given on how the resource balance is oriented: towards the use (and even storage) of nutrients from the acquired food, or towards the mobilisation of endogenous reserves.

With the aid of diagnostic tools similar to those used in human and veterinary medicine, body fluid biochemistry can be used to assess body condition and individual health in other organisms (Resano-Mayor et al., 2016; Schoepf et al., 2017), especially for micro-nutrients only acquired by food. Since the late 1990s, interest in non-lethal methods to evaluate the physiological state of individuals has grown and has stimulated the development of technical devices and commercial kits compatible with the measurement of micro-volumes. Thus, biochemical analyses of plasma/serum or hemolymph can be conducted to measure various markers related to nutrition and digestion, metabolism, nutrient storage, osmoregulation, liver or kidney function, and immunity. Specifically, nutritional status and patterns of energy utilisation can be assessed by measuring the levels of the products of intermediary metabolism related to diet and detectable in body fluids. This is the case for total proteins, glucose, albumin, urea, creatinine, uric acid, essential minerals, and trace elements. Lipid metabolism can be assessed by measuring the concentrations of lipids and ketones, homeostasis, energetic metabolism, or the functioning of tissues involved in digestion based on the activities of specific enzymes (e.g. alkaline phosphatase, lactate dehydrogenase and creatine kinase). There are also markers specifically associated with starvation, such as bile acids, bilirubin, and globulin, and those that allow accurate assessments of hepatic, renal, and pancreatic function, or electrolyte levels (major and dietary minerals). Analyses of body fluids may also target specific micronutrients essential for normal metabolic processes and thus potentially indicative of health status. An example is carotenoids. These fat-soluble pigments originate from plants, algae, or fungi but cannot be synthesized de novo by vertebrates, which must acquire them exclusively from dietary sources. Hence, carotenoid-based traits can reveal the foraging ability, resource availability, and trophic interactions, as well as the nutritional and immune status of their bearers (Sternalski et al., 2010).
The main drawback of these techniques lies in the difficulty of interpreting the data obtained from the battery of markers. The measured parameters must be chosen carefully to obtain convincing evidence of a metabolic pattern, or change in nutritional status. Furthermore, some parameters can be interpreted only in the light of variations in others (for instance lipids or hepatic enzymes), and body fluid biochemistry can be affected by several confounding factors in the relationship with food acquisition (for instance, blood/hemolymph metabolite concentrations may change after food ingestion, and age, sex, and reproductive stage can affect metabolic levels and enzymatic activities). Given these considerations, an integrated assessment is critical. It should be based on the use of several body fluid biomarkers or other metrics of body condition and should take into account individual and ecological factors as well (Resano-Mayor et al., 2016; Wilder et al., 2016). Indeed, changes in physiological conditions are not always food-related but may be driven by other abiotic stressors (e.g. temperature changes, contaminants). Such stressors might be considered carefully because they could be used to get further insights into the consequences of the interactions between environmental factors and trophic relationships (see for instance Bustnes et al., 2015). Such issues are raising growing interest within the framework of stress ecology. Nutritional ecology is one side within trophic ecology framework that is perhaps not enough considered to build integrated approaches and may deserve further attention in studies on food webs and trophic ecology (Raubenheimer et al., 2009).

4.2. Trait-based trophic ecology

Focusing on functional features rather than on taxa helps describing biodiversity from a functional point of view (Levine, 2016). Trait-based approaches can be an adequate theoretical framework for understanding and predicting food webs dynamics and properties (Bartomeus et al., 2016; Gravel et al., 2016). Thus, trait-based trophic ecology investigates both (1) trophic interactions per se and (2) their ecosystem implications. First, functional traits allow inferences regarding the most probable interactions occurring within a community. A common first approach uses phenological traits or ecological preferences to detect species that co-occur and to discard unlikely trophic interactions (González-Varo and Traveset, 2016). The morphological and/or physiological traits of the co-occurring partners are then used to infer the probability of realised trophic interactions (see Gravel et al., 2016 for a useful framework distinguishing foraging, vulnerability and consumption traits). Because determining all interactions among individuals, even in simple systems, is fairly unattainable, narrowing those interactions using functional traits provides an alternative and can make the mechanistic underpinnings of interactions understandable (e.g. Ibanez et al., 2013). In addition, functional traits enable the different dimensions of the trophic niche to be investigated individually or simultaneously by examining the morphological, physiological, phenological and behavioural determinants of the trophic niche of a particular species (e.g. Spitz et al., 2014).

Two tenets of functional-trait-based approaches are essential in interpreting the implications of the identified trophic interactions. First, ‘response and effect traits’ link the response of individuals to higher trophic levels or to environmental stressors with the potential effects on lower trophic levels. For instance, plant diversity changes affect functional diversity of arthropods across trophic levels (Ebeling et al., 2018) which might modulate ecosystem processes. Second, functional-trait-based approaches can explicitly address intra-specific variability at the individual level (Albert et al., 2011; Rota et al., 2018) and the findings then scaled up to the community and ecosystem levels (e.g. Lavorel and Grugulis, 2012).

Despite recent progress (e.g. methodological standardization Moretti et al., 2017), the development of functional-trait-based approaches in trophic ecology is still hampered by several limitations and drawbacks. For example, it remains difficult to assess the strength of species interactions (the direct effect that species have on their mutual demography). Nevertheless, it is possible to estimate functional food web relationships by distinguishing (1) the life-history traits which may modulate the strength of an interaction on population growth, from (2) the topological traits which may modulate the pairwise interactions between the predator and its prey, and from (3) the consumption traits which may modulate the per capita interaction strength (Gravel et al., 2016, see also Brousseau et al., 2018). Additionally, the correlation between individual traits and how this lack of independence among traits reflects phylogenetic (evolutionary) constraints remain poorly understood (Poff et al., 2006). Thus, the validity of linking traits to actual trophic functions continues to be a source of debate, as the causal link between them is often far from obvious, quite variable, and/or has yet to be definitively demonstrated.

4.3. Ecological stoichiometry (ES)

The consequences of elemental imbalances in ecosystems can be explored using ES approaches, in which the elemental compositions of heterotrophic organisms (generally expressed as C:N or C:P ratios) are considered as far less variable than those of primary producers, at least for most similarly sized taxa (Persson et al., 2010). Based on the law of conservation of matter, ES approaches can predict how organisms are influenced by the mismatch between consumer demand and the elemental supply available in resources (Sterner and Elser, 2002). At the ecosystem scale, measuring imbalances can be used to estimate nutrient transfer efficiency and secondary production (Cebrian et al., 2009). In the presence of excess elements in their food, consumers make use of several coping strategies such as: (1) selecting food with elemental ratios close to their requirements and/or (2) eating food of different qualities but assimilating only those elements that fulfill their requirements while rejecting those available in excess. In the latter case, mass balance models can estimate the amount of excess elements released. Experimental measurements of metazoan’s nitrogen and phosphorus excretion rates showed that animals can supply nutrients at rates similar to those of other major nutrient sources and support a large portion of primary production by nutrient recycling (Vanni, 2002; Vanni et al., 2002). Excreted elements reduce nutrient limitations and impact ecosystem processes in a predictable manner (Dangor et al., 2009). Accordingly, ES is a powerful conceptual framework for predicting both the consequences of resource ingestion on consumers’ life history traits and the ecosystem consequences of elemental imbalances. To date, most ES studies have focused on C, N and P; however, the consideration of other essential elements (e.g. As or Cu, see Karimi and Folt, 2006) might provide a better understanding of nutrient transfer and the potential limitations for consumer growth or ecological processes (Welti et al., 2017).

4.4. Resource quality

Parameters other than the C:N:P ratios of resources can influence consumers’ growth and physiological status as well as the efficiency of nutrient and energy transfer into food webs. These include several essential compounds as crucial drivers of diet selection (Kohl et al., 2015). For example, the lack of certain polyunsaturated FAs of the ω3 series and/or phytosterols limits Daphnia growth and/or reproduction (e.g. Martin-Creuzburg et al., 2009). Micronutrients are another example and include carotenoids, described in Section 4.1.2. The pigments are important stimulators of immune responses and show antioxidant properties (e.g. Lucas et al., 2014). Integrative approaches that include large classes of biochemical compounds (proteins, lipids, glucids) have been proposed to relate resource composition and organism fitness within a conceptual framework referred to as “nutritional geometry” (Simpson and Raubenheimer, 1993). Its advantage is that it directly relates consumers’ feeding strategies to their life history traits (e.g. optimal carbohydrate/protein ratios that optimize either individual growth or reproduction) and thus nutritional geometry allows
evolutionary questions to be explicitly related to trophic ecology. Moreover, a combination of all of these resource-quality-based approaches has been proposed (Sperfeld et al., 2017) and is a promising methodological strategy for research in trophic ecology.

5. Testing the implications of trophic interactions (trophic models)

While inference methods are used to reconstruct trophic interactions within community (Section 3), modelling approaches presented in this section allow investigations of the consequences of feeding interactions on various ecosystem features, ranging from population to ecosystem dynamics. The assumptions underlying the different models include continuous/discrete, mechanistic/statistical and individual—/population-based systems. The common goal is to simulate food web functioning from a set of given conditions, such as network structure, which can in some cases be determined through inference methods. Modelling approaches can be used to investigate the potential implications of feeding interactions in terms of community dynamics and/or organic matter/energy transfers (e.g. Rall et al., 2008) and thus contribute to an improved understanding and management of ecosystems. The use of models also makes possible to investigate hypotheses that cannot be tested in natural systems (e.g. Gaucherel et al., 2017), as well as predictions regarding ecosystem responses to management decisions (e.g. Marzluff et al., 2016). In this section, we present the main modelling approaches currently used for hypotheses testing in trophic ecology.

5.1. Ordinary differential equations and qualitative models

Ordinary differential equations (ODEs) simulating species dynamics, such as the Lotka-Volterra equations, can be adapted for food web models. The latter have been central to understanding indirect interactions in food webs and in studying the consequences of predator-prey interactions on population dynamics (e.g. Rall et al., 2008). Food web models based on ODEs allow investigations of the relationship between food web complexity and stability (e.g. McCann and Rooney, 2009). During the last two decades, models based on ODEs have been expanded to include allometric constraints on species traits, such as metabolic rates and feeding rates, leading to the so-called bioenergetic or allometric trophic network model (Brose et al., 2017). Ecosim, the time dynamic simulation module associated with Ecopath, is a well-known application of this approach and it is commonly used to explore policy options in the ecosystem-based management of fisheries, such as the effects of fishing on ecosystems and the establishment of marine protected areas (see Walters et al., 1997).

Qualitative modelling focuses on the feedback structure of food webs and is used to assess both stability and ecosystem responses to perturbations (Puccia and Levins, 1985). This method, which is also based on ODEs, is particularly well suited for trophic ecology studies, as it does not rely on the quantification of all trophic interactions within a system. Indeed, based only on qualitative assessments of the interactions among major trophic compartments (e.g. positive, neutral or negative), qualitative models of food webs can provide a general understanding of ecosystem stability and dynamics based on network topology (Halof et al., 2000). For instance, they can be used to predict the overall responses of a community to long-term disturbances, as the spatial shift of multiple marine species due to ocean warming (e.g. Marzluff et al., 2016). However, the predictions are most useful for simple networks (<15 nodes) as they become ambiguous in more complex systems (e.g. Dambacher et al., 2003).

5.2. Statistical approaches

Statistical approaches are based on graphical models (see Section 3.1), but in the present case, the graph structure (i.e. the trophic network) is known and is not an output of the model. Trophic interactions are represented as edges in the graph and are interpreted as statistical interactions between species by the model. Using the resulting graph, statistical approaches are able to model the probability of species persistence, explicitly, without a mechanistic model of the extinction dynamics. Unlike Lotka-Volterra or structural food-web models that are based on material fluxes, statistical approaches can be built from simple in situ observations of individual occurrence. The required experimental set-up is thus simpler than the one for computing the parameters and structure of Lotka-Volterra or structural models. Another advantage is the possibility to evaluate the resilience of trophic networks using the probabilities of species persistence, and not only the trophic structure. McDonald-Madden et al. (2016) adopted this approach to study the conservation management of 6 real food webs (the Alaskan, Baltic Sea, Lake Vattern, Chesapeake Bay, Arizona Mountain forest and Long Island Salt Marsh food webs) and 40 hypothetical food webs. The resulting predictions allow estimating the effect of management strategy directly at the scale of food-web and their study provide a computationally efficient way of identifying the important species to manage in large food webs.

5.3. Discrete approaches

In discrete approaches, networks of trophic interactions are represented by systems made of distinct and separate components, and characterised by temporally distinct and separated events (e.g. Campbell et al., 2011). As such, discrete approaches, more than classical trophic models, are particularly appropriate to consider complex interaction networks (i.e. networks with a high number of direct and indirect processes), while their behaviour remain under rigorous control. A strong advantage of discrete models is thus the possibility to model the evolution of trophic networks over time. Studying ecosystem trajectories constitutes a potential fruitful research avenue for discrete approaches that could help identifying sustainable paths (the ecosystem is able to persist and evolve) from dead-ends (the ecosystem disappears or is stuck in a specific state with less interacting species). Gaucherel et al. (2017) followed such an approach, using Boolean networks, to study the trajectories of a theoretical termite colony. By removing different numbers and types of nodes, the authors simulated different disturbances of the colony and examine the effects on the interaction network. By doing so, they showed that the colony was highly sensitive to the trophic part of the interaction network. In addition, this example illustrates the conceptual shift permitted by discrete approaches towards broader (spatial and temporal) scales of ecological processes (Wallach et al., 2017).

5.4. Individual-based models

In an individual-based model (IBM), each individual and its associated behaviour are represented, which allows interactions among individuals to be approximated. IBMs are commonly used in ecology (Grimm et al., 2005). An important research activity surrounding IBMs consists in understanding how patterns at the population or community scale emerge from the interactions among individuals (Durrett and Levin, 1994). In trophic ecology, IBMs can be used to reveal the consequences of intraspecific variations in diet on community trophic structure (Bolnick et al., 2011) or eco-evolutionary dynamics (Melian et al., 2011). Another application consists in scaling from trait-based interactions between individuals to the global structure of ecological networks (Jabot and Bascompte, 2012).

5.5. Dynamic energy and mass budget approaches

The dynamic energy and mass budget (DEB) theory describes the organisation of metabolism within organisms (Sousa et al., 2008). The ground principle consists in modelling the quantitative facets of metabolism (assimilation rate, growth/maintenance, and maturity/reproduction) using common rules of allocation and the processes occurring at the organism level. Standard DEB model is a system of three
ordinary differential equations following first order dynamics. Assimilated products first enter a reserve pool which is then mobilized to fuel two pathways following the kappa rule: a fixed kappa fraction is allocated to perform growth and maintenance and the remaining fraction (1-kappa) is available for maturity (i.e. increase of complexity and its maintenance) and reproduction (Van Der Meer, 2006). A set of 12 primary parameters is calibrated from life history traits and univariate data (Lika et al., 2011). Common model inputs are temperature and food availability and outputs are size, shape, full life cycle dynamics, reproduction rate, and composition of the organism (C, H, O, N). Standard DEB models (one reserve, isometric organism) are commonly used at the level of the individual organism (e.g. Grangeré et al., 2010) but they can be scaled up to population level (Martin et al., 2012) or to food web and ecosystem levels including trophic interactions (e.g. Marques et al., 2014). Interesting applications of DEB models in trophic ecology are related to diet reconstructions (Picoche et al., 2014), the effect of food stoichiometry on growth (Muller et al., 2001), the effect of trophic ecology is a vibrant field of research supported by a diverse and ever-growing array of methodological tools. For ecologists seeking to answer specific trophic questions, we examine this set of tools (Table 1) based on our own experience and background (Fig. 1). For the empirical methods, our discussion includes the results of a multiple correspondence analysis (MCA) based on an evaluation of the price, investment, and affinities of each method with key criteria, such as the observational scale, organisational level, type of ecological question to be tackled, and the type of data produced (Fig. 1A; the procedure is detailed in the Appendix). The first axis of the MCA was mainly driven by the type of ecological question and the second axis was related to the investment needed to implement the methods. On the same figure (Fig. 1A), we also displayed the methods’ affinity with organisational level and a rough evaluation of their costs. We completed this analysis with a subjective classification of the various trophic models (after C. Gaucherel and E. Thébault, pers. com.). On the resulting Fig. 1B, we focused on the models’ affinity with the type of ecological question and organisational level for the sake of consistency with the classification of empirical methods. Not included into this classification were inference methods, because they could not be discriminated by these two axes. The resulting roadmap (Fig. 1A and B) provides an overview of the affinities of the methods for the different criteria. As an example, from Fig. 1A it can be deduced that the use of observational methods is more appropriate than stoichiometry if the nature of feeding interactions is the subject of interest. This visual overview also highlights the gaps (i.e. the absence of available method to address a specific issue), such as the modelling of fluxes at the integrated scale of ecosystems (and not only between species, see Fig. 1B). An advantage of the roadmap is that it facilitates the selection of the best combination of methods with which to investigate (or triangulate) a specific issue in trophic ecology (see below).

7. Interconnecting approaches to advance ecological research

The abundance of methods currently available to trophic ecologists offers opportunities such as the triangulation referred to by Munafò and Smith (2018), i.e. the intentional use of multiple approaches which complementarity offers less-biased, multiple lines of evidence. In other words, combining two or more methods can overcome the shortfalls of each one alone, and initiate interdisciplinary collaborations that will enrich our knowledge and perspectives. More than combining several methods at the same integration level (e.g. SIA + FA + Gut contents), we recommend more interconnection of approaches, for example by creating first a topology using traits, then modelling and using ingestion/assimilation tracers as validations. Only a few comparative studies of methodologies have been published (but see Nielsen et al., 2018), in the following we present several examples in which complementary methods were merged or interconnected and led to novel insights.

Combinations of methods have been used to improve empirical descriptions of food webs, especially as there are recurring issues related to the integration of trophic processes over time. These can be addressed by complementary approaches. For example, while assimilation tracers such as isotopes and FA s are regularly combined, the addition of complementary behavioural observations and ingestion tracers would favour the emergence of comprehensive theories and models describing tropho-dynamics in food webs (e.g. Woodward et al., 2010; McMeans et al., 2016). Direct field records (motion-sensitive cameras, Robinson et al., 2018) are also useful additional measurements to obtain inferred food webs with a much better temporal accuracy. At the individual level, SIA s can be coupled to DNA-based analyses of gut contents to enable the detection of possible cannibalistic behaviour, as this is not possible with molecular methods (Traugott et al., 2013), and to overcome the lack of dietary resolution inherent to stable isotope methods (e.g. Hambäck et al., 2016).

Other examples show that interconnecting trait-based concepts with stable isotope methods sheds light on the interplay between interactions and fluxes. In fact, this has contributed to the emergence of the concept of isotopic trophic niches (Bowes et al., 2017, and references therein). Also, replacing predator-prey functional traits (e.g. predator biting force vs prey cuticle toughness) within a phylogenetic framework (e.g. evolutionary distance between trophic levels) can give rise to models of predator-prey interactions (Brousseau et al., 2018). Observational data can contribute to inference methods and models to obtain a characterisation of the ecosystem-level properties emerging from trophic interactions (e.g. Pacella et al., 2013; De Vries et al., 2013). For example, a Bayesian model using topology and isotopes was specifically dedicated for the elucidation of trophic strengths in trophic networks (IsoWeb, Kadoya et al., 2012). Models tested against empirical data can be employed to reveal the mechanisms operating within food webs and to re-assess the relevant level of complexity (guilds) at which food webs should be scrutinised in the field (Halot et al., 2000). Theoretical modelling and empirical data, connected through a trait-based approach to species and metabolic rates within allometric trophic networks, can successfully reproduce complex multi-species dynamics in natural environments (Boit et al., 2012). Among the most recent attempts involve hybrid modelling, in which models of the overall dynamics of the ecosystem are combined with realistic individual-based models to link responses at the individual and population levels (Strauss et al., 2017). Also worthy of mention are stoichiometry-based approaches, which offer a powerful and promising perspective from which to extend our knowledge about food webs to include biogeochemical consequences and effects on ecosystem services (Boit et al., 2012; Strauss et al., 2017). A stoichiometric-trait-based approach was recently described (Meunier et al., 2017), although it requires further development (Welti et al., 2017). Multivariate DEB models (several substrates, or/and reserves and structures) offer new perspectives to integrate nutritional geometry and ecological stoichiometry into a metabolic theory through the concept of organisms’ homeostasis (Sperfeld et al., 2017). Capturing complex trophic interactions within or between species could then be modelled by embedding multivariate DEB models in agent-based modelling approaches (Sperfeld et al., 2017). Finally, DEB models estimate key morphological traits, such as size, weight, shape, ingestion rates, and range of temperature tolerance. By doing so, they provide connections to biophysical models (e.g. temperature, wind speed and radiation) and to nutritional geometry that make possible to model trophic and ecological niches of organisms ( Kearney et al., 2010).
Table 1
Synthesis of the main advantages and drawbacks of several methods commonly used in trophic ecology. Expert assessment of empirical methods according to 12 variables is also shown: price, investment (is it a turnkey method? or does it require further development?), observational scale (from microscopic to large organisms), organisational level (from individual to ecosystem), type of ecological question (interaction vs. flux of energy/matter), and type of produced data (qualitative vs. quantitative); 0 indicates “not adapted” and 1 indicates “strong affinity”. The exceptions are the variables “price” and “investment”, where 0 and 1 stand for “low” and “high” respectively, and the “data” variable, where 0 and 1 stand for “qualitative” and “quantitative” data respectively.

<table>
<thead>
<tr>
<th>Detecting prey-predator relationships, or food sources</th>
<th>Price</th>
<th>Investment</th>
<th>Micro</th>
<th>Small</th>
<th>Large</th>
<th>Individual</th>
<th>Population</th>
<th>Community</th>
<th>Ecosystem</th>
<th>Interaction</th>
<th>Flux</th>
<th>Data</th>
<th>Advantages</th>
<th>Drawbacks</th>
</tr>
</thead>
<tbody>
<tr>
<td>Observation of feeding activity</td>
<td>0 1 0</td>
<td>1 1 1 1 1 0</td>
<td>0 1 0</td>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Straightforward and well-established / Relatively cheap / Various interactions and scales (incl. intraspecific variability) / In situ or under controlled conditions / Large scale records of feeding-behaviors are possible with miniaturization of loggers</td>
<td>Temporal &amp; spatial scales are design-dependent/ Difficult to measure assimilation</td>
</tr>
<tr>
<td>Incubation experiments</td>
<td>0 1 1</td>
<td>1 1 1 0 1 0 1</td>
<td>1 1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Effect-size of specific variables / Suitable for filter-feeders</td>
<td>Mostly laboratory-based, so deviation from natural conditions might occur</td>
</tr>
<tr>
<td>Gut &amp; feces content</td>
<td>0 1 1</td>
<td>1 1 1 1 1 0</td>
<td>0 1 0</td>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Well-established / Relatively cheap / Information on nature and size of ingested food items</td>
<td>Snapshot of diet (does not reveal interaction strength) / Time consuming / Labile tissues or prey might be overlooked</td>
</tr>
<tr>
<td>Gut DNA</td>
<td>1 1 0</td>
<td>1 1 1 1 1 0</td>
<td>1 0 1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Identification of gut microbiomes / Works on partly digested, poorly recognizable, food items</td>
<td>Expensive / Susceptible to typical drawbacks associated to molecular approaches</td>
</tr>
<tr>
<td>Parasites as biological tags</td>
<td>0 1 0</td>
<td>0 1 1 1 1 0 1 0</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Host specificity / Integration of diet over long time periods / Potential information on diet provenance</td>
<td>Lethal / No information on prey size / Needs empirical knowledge of life-cycles (and potentially biogeography) of parasites / Bias due to side-effects of parasites on hosts’ health and behavior</td>
</tr>
<tr>
<td>Bulk Stable Isotope Analysis</td>
<td>0 0 0</td>
<td>1 1 1 1 1 1 0 1 1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Well-established / Relatively cheap / Traces the C and N food sources in consumers / Important replication is possible / suited for in situ as well as laboratory studies</td>
<td>Turnover times and isotopic discrimination might vary among species, tissues and the environment studied / Not well-suited when a large number of potential resources are considered</td>
</tr>
<tr>
<td>Isotopes as labels</td>
<td>1 1 0</td>
<td>1 1 1 1 1 1 0 1 1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Trace the fate (timing and scale) of carbon and nitrogen from microbes to macroscopic consumers / Differentiation between N and C uptake from unlabelled C and N pools</td>
<td>Difficult to provide quantitative fluxes / Needs to know turnover incorporation and routing in tissues (e.g. issue of uniformly labeling the food source) / Can be relatively expensive depending on the scale or nature of the labeling</td>
</tr>
<tr>
<td>Fatty Acids</td>
<td>1 1 1</td>
<td>1 1 1 1 1 0 1 0</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Well-established / Traces flux and nutritional quality</td>
<td>Non-specific biomarkers / Needs a priori knowledge of producers’ and consumers’ metabolism / relatively expensive</td>
</tr>
<tr>
<td>Compound-Specific Isotopic Analysis</td>
<td>1 1 0</td>
<td>1 1 1 1 1 1 0 0 1 1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Integrative tracers when coupled with a labeling / Target analysis promising in Microbial Ecology / Ideally it can trace limiting compounds and eventually detect minor food sources unseen by classical methods / Better knowledge in fractionation and turnover times for specific compounds</td>
<td>Needs a good understanding of the biochemical and physiological pathways in investigated organisms or tissues / Relatively expensive</td>
</tr>
<tr>
<td>Contaminants</td>
<td>1 1 1</td>
<td>1 1 1 1 1 1 0 1 1 0</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Might trace spatio-temporal ingestion and assimilation depending on the persistence of compounds within tissues (e.g. lipophilic contaminants might biomagnify along food chains) / Potentially information on foraging area, and on contamination status of species and ecosystems</td>
<td>Needs a source of contamination / Biases due to side-effects on health and behavior / Needs knowledge of contaminants’ biogeochemical cycle, distribution and persistence in organisms and in the environment / Relatively expensive</td>
</tr>
<tr>
<td>Connecting species within networks from ecological data (inference methods)</td>
<td>Advantages</td>
<td>Drawbacks</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>---</td>
<td>---</td>
<td>---</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>Gaussian graphical models</strong></td>
<td>Popular framework to infer interactions among continuous variables / Obtained precision matrix is a direct proxy of network structure</td>
<td>The network is undirected (unlike trophic/parasitic relations) / High computational complexity (sparse matrix inversion) / Unsuitable for classes-occurrence observations / Assume a gaussian distribution of interactions</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>Bayesian networks &amp; dynamic Bayesian networks</strong></td>
<td>Adapted to class-occurrence data / Suitable to represent directed interactions (trophic, parasitic...) / Can be used to infer ecosystem static states or dynamics / Numerous heuristics are available to reach efficient approximate inference</td>
<td>Not adapted to continuous observations / Exact inference is out of reach given the combinatorial nature of the underlying optimization problem. / The learnt network describes &quot;deterministic&quot; interactions / Accuracy of learning is hard to validate because the learning process is rather &quot;black box&quot;</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>Logic-based approaches</strong></td>
<td>Adapted to qualitative data / Can incorporate background (expert) knowledge on the interactions / Describes directed interactions / Requires relatively less data than others</td>
<td>Not adapted to continuous observations / Exact inference is out of reach given the combinatorial nature of the underlying optimization problem. / The learnt network describes &quot;deterministic&quot; interactions / Accuracy of learning is hard to validate because the learning process is rather &quot;black box&quot;</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>Linear inverse inference</strong></td>
<td>Particularly suited to infer the magnitude of missing interactions based on simple assumptions</td>
<td>Relies on a known network topology and on the static assumption of mass balance between all network compartments at equilibrium.</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>Approximate Bayesian computation</strong></td>
<td>Replaces the step of likelihood computation by simulations of the model with variable parameter values</td>
<td>Requires substantial computing time</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Quantifying the consequences of feeding from individuals to ecosystems</th>
<th>Price</th>
<th>Investment Micro</th>
<th>Small</th>
<th>Large</th>
<th>Individual</th>
<th>Population</th>
<th>Community</th>
<th>Ecosystem</th>
<th>Interaction</th>
<th>Flux</th>
<th>Data</th>
<th>Advantages</th>
<th>Drawbacks</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Body composition</strong></td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>Measuring energy storage is one of the most direct and quantitative indicator of body condition</td>
<td>Dissection might be time-consuming and tedious / Alternative techniques are less invasive but more expensive and require further development</td>
</tr>
<tr>
<td><strong>Body condition indices</strong></td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>Integrative assessment of physiological state (health, diet deficiencies)</td>
<td>Physiological state can be affected by other factors than nutritional stress / Difficult interpretation that needs integrated assessment (combination of markers) / Expensive / Less suitable on small animals</td>
</tr>
<tr>
<td><strong>Body fluid biochemistry</strong></td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>Integrative assessment of physiological state (health, diet deficiencies)</td>
<td>Physiological state can be affected by other factors than nutritional stress / Difficult interpretation that needs integrated assessment (combination of markers) / Expensive / Less suitable on small animals</td>
</tr>
<tr>
<td><strong>Traits</strong></td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>1</td>
<td>0</td>
<td>Essential compounds are key drivers of diet selection along the life of an organism / Existence of a &quot;nutritional geometry&quot; framework</td>
<td>Young field of research / Needs integrative, and thus potentially expensive, measures</td>
</tr>
<tr>
<td><strong>Ecological Stoichiometry</strong></td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>Essential compounds are key drivers of diet selection along the life of an organism / Existence of a &quot;nutritional geometry&quot; framework</td>
<td>Young field of research / Needs integrative, and thus potentially expensive, measures</td>
</tr>
<tr>
<td><strong>Resource quality</strong></td>
<td>1</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>Essential compounds are key drivers of diet selection along the life of an organism / Existence of a &quot;nutritional geometry&quot; framework</td>
<td>Young field of research / Needs integrative, and thus potentially expensive, measures</td>
</tr>
</tbody>
</table>

**Table notes:**
- **Price:** 1 = High, 2 = Medium, 3 = Low
- **Investment:** 1 = High, 2 = Medium, 3 = Low
- **Micro:** 1 = High, 2 = Medium, 3 = Low
- **Small:** 1 = High, 2 = Medium, 3 = Low
- **Large:** 1 = High, 2 = Medium, 3 = Low
- **Individual:** 1 = High, 2 = Medium, 3 = Low
- **Population:** 1 = High, 2 = Medium, 3 = Low
- **Community:** 1 = High, 2 = Medium, 3 = Low
- **Ecosystem:** 1 = High, 2 = Medium, 3 = Low
- **Interaction:** 1 = High, 2 = Medium, 3 = Low
- **Flux:** 1 = High, 2 = Medium, 3 = Low
- **Data:** 1 = High, 2 = Medium, 3 = Low
Testing the implications of trophic interactions (trophic models) | Advantages | Drawbacks
--- | --- | ---
Ordinary Differential Equations and qualitative models | Understanding of indirect interactions and species dynamics in food webs / Qualitative models useful when the quantification of each trophic interaction is unreachable / Useful to investigate potential consequences of perturbations on food web functioning and stability | ODE models for large food webs often require a disproportionate number of parameters, and all parameter combinations are impossible to test through sensitivity analysis / Predictions of the effects of perturbations with qualitative models become ambiguous for large networks (< 15 nodes)
Statistical approaches | Can be built from simple field observations of individual occurrences / Useful to evaluate management strategies dedicated to increase food web resilience (operational models) | Not process-based, thus no mechanistic understanding / Dependant on collected data (over- and under-fitting)
Discrete approaches | Adapted to discrete and qualitative interactions / Well adapted to large interaction networks, whatever the nature of components and processes / No need of calibration, except the expert knowledge defining the model / Relevant to deduce all possible trajectories and stabilities the system may experience. | Hardly adapted to continuous quantitative abundances, biomasses or fluxes in the network
Individual-based models | Help understanding how patterns emerge from inter-individual interactions / Able to scale up from trait-based interactions to structure of ecological networks | Often require very intensive numerical simulations (calibration and validation stages) / Not adapted to heavy individual or multiscale networks / Hardly provide rigorous and demonstrated analyses
Dynamic energy and mass budget approaches | Model the quantitative facets of metabolism by depicting rules of allocations and processes at the organism level / Application to diet reconstructions | Huge personal investment required / Decreasing interest when used for complex applications at community or ecosystem levels

8. Including the microbial world in trophic ecology

The microbial world has long been treated almost independently from the macroscopic world (Prosser et al., 2007). Conversely, studies of microbial diversity do not explicitly consider the concepts of trophic ecology. The emergence of molecular approaches has enabled microbial cryptic diversity to be revealed through genomics, by mapping co-occurrence networks within microbial communities (Barberán et al., 2012; Peura et al., 2015). However, the implications of the results in terms of trophic interactions within microbial communities or in relation to the metazoan food web remain poorly explored (Peura et al., 2015). On the other hand, trophic ecology has not yet explicitly integrated the role and contribution of complex microbial communities even though the participation of microbes in aquatic and marine food webs was described more than four decades ago (Pomeroy, 1974), and heterotrophic protists have been recognized as the major grazers in the oceans (Calbet and Landry, 2004; Sherr and Sherr, 2007). Bacterial and micro-eukaryotic communities are, for the most part, included in food web models as broad, unresolved taxonomic, or functional groups, with very few exceptions (Peura et al., 2015; D’Alelio et al., 2016; Weitere et al., 2018). Furthermore, estimates of trophic chain length fail to consider heterotrophic microbes (Post, 2002), while recent CSIA-based assessments register heterotrophic microbes in food webs at the same trophic positions as animals (Steffan et al., 2015).

Consequently, our understanding of the “green” and “brown” food webs, including the tropho-dynamic processes of the detritusphere, emphasised already by Lindeman (1942), remains deficient, and the potential contribution of the topology of microbial and metazoan networks to ecosystem stability and resilience barely known (Peura et al., 2015). Several recent methodological developments may be the first steps in ground-breaking advances in our understanding of micro-macro food webs: (1) Recent developments in DNA-SIP (Pepe-Ramney et al., 2016) and RNA-SIP (Kramer et al., 2016) allow the tracking and identification of microbial metabolisers of organic substrates up to primary consumers (see Section 2.4.3), thereby improving empirical descriptions of the links within prokaryotes and protists but also allowing a quantification of the connection to higher trophic levels. (2) By taking advantage of the surge of metagenomic and metatranscriptomic approaches, microbial ecologists have been able to link microbial genetic diversity with the functionalities and biogeo-chemical processes determined in field samples (see Morales and Holben, 2011 and references therein). (3) Information on taxon-specific feeding selection and the rates measured in experimental settings from microbial and metazoan organisms can be used as input in Ecopath-like food web models adapted to explicitly include highly resolved protozoan contributors (D’Alelio et al., 2016). These models can then be applied to explore the functional consequences of the switches in microbial food web structure on the overall trophic transfer of matter across the micro-macro continuum (D’Alelio et al., 2016). Ecological stoichiometry (Welti et al., 2017) or the application of trait-based approaches to microorganisms (Litchman et al., 2015) may serve as a bridge between the microbial food web and the energetics and biogeochemistry of the ecosystem, while allometric scaling could connect processes at the food web scale, thus merging the “green” and “brown” food chains (e.g. Mulder et al., 2013).

For trophic ecologists, these elements well evidence the so-far-unrealised opportunity to fully consider vertical biodiversity based on an explicit representation of the micro- and macroscopic actors and their roles. With the improvement of the collaboration between microbiologists and trophic ecologists many long-standing questions will finally be answered and “complexity made simple” (Pomeroy, 2001).

9. Conclusion

In a world of fast and dynamical changes in environmental conditions, there is a crucial need to understand and anticipate changes in biodiversity in both space and time. Changes in abiotic factors have direct effects on species that depend on specific tolerance, physiological acclimation, phenotypic plasticity, and evolutionary processes (Angert et al., 2013), which likely create local changes of biodiversity, such as spatial redistribution of species (Marzloff et al., 2016). Yet, evidences are piling that indirect effects of global changes through alterations in species interactions overweight direct effects (see Holt and Barfield, 2009; Angert et al., 2013) or that species range shifting can induce new trophic cascades, inflating the ecological consequences of multiple species redistribution (Marzloff et al., 2016). Such observations of the
crucial role of feeding interactions in the biodiversity response to global changes place trophic ecology at the core of the issue (see R. Holt contribution to Nature 2020 visions, Norvig et al., 2010). Recent conceptual and methodological developments in trophic ecology now make possible to explore all the dimensions of such changes, but also to draw cross-scale linkages, at least for some pilot sites and case studies. Numerical improvements and machine-learning technics offer the opportunity to test theories and capitalize on these laboriously obtained field- or lab-data by expanding the time- and space scales of processes and predictions (Faisal et al., 2010; Angert et al., 2013). We hope this synthesis will convince our colleagues that we have at hand the perfect toolbox to finally deal with the enormous complexity of trophic implications.

**Conflict of interest**

The authors have no competing interests to declare.

**Acknowledgements**

This contribution is a collective effort by the French network for trophic ecology (GRET: Groupe de Recherche en Ecologie Trophique), supported by CNRS, INEE and INRA as GDR 3716. We are grateful to the numerous members of the GRET who attended workshops in Toulouse and Nantes, where fruitful discussions led to this review. We address many thanks to Craig Layman for his helpful comments on a previous draft of our manuscript. MEP proposed this review and NM, NHT and MEP led, organized and completed the writing. CG provided helpful support for Sections 3 and 5, and ET, AB and MD for Section 7. All authors contributed critically to the drafts and gave final approval of the manuscript for publication.

**Appendix A. Supplementary data**

Supplementary data to this article can be found online at [https://doi.org/10.1016/j.jfooweb.2018.e00100](https://doi.org/10.1016/j.jfooweb.2018.e00100).

**References**


**Fig. 1.** (A) Multiple correspondence analysis showing the general distribution of empirical methods used in trophic ecology (see Table 1 for data and abbreviations). Arrows indicate the affinity of the methods with the selected features: organisational level (from individuals to ecosystems), ecological questions (interaction vs. flux of energy/matter), or commitment of budget, time, and efforts (investment, price). Observational scale (i.e. microscopic to macroscopic) was mostly discriminated by axis 4, not shown here, but scores are in appendix. (B) Subjective classification of trophic models based on their affinity with the organisational level and ecological question (after C. Gauchereel and E. Thébault, pers. com.). ODE: ordinary differential equation, IBM: individual-based model, DEB: dynamic energy and mass budget.


Pomeroy, L.R., 1974. The ocean’s food web, a changing paradigm. Bioscience 24, 499–504.


